



ASSOCIATION OF **RINGSIDE PHYSICIANS**

Blood-borne Infection Prevention in Combat Sports

Position Statement of the Association of Ringside Physicians

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POSITION STATEMENT (ABSTRACT)

The Association of Ringside Physicians (ARP) emphasizes the importance of screening combat sports athletes for blood-borne infections, including hepatitis B, HIV, and Hepatitis C, to mitigate transmission risks and ensure participant safety. Although transmission of hepatitis B and C and HIV in combat sports are rare, protecting athletes is of utmost importance. It is the recommendation of the ARP that all fighters participating in combat sports in which the presence of blood is a common occurrence and is allowed during competition, should undergo testing for HIV, Hepatitis B (HBV), and Hepatitis C (HCV). Testing should be conducted using serum samples, as rapid tests are not considered acceptable for accurate results. Testing for HBV, HCV and HIV should optimally be done within 3 months of competition, but within 6 months is acceptable. Athletes whose tests suggest active HBV, HCV or HIV infection should be disqualified from competition in sports where blood is common and allowed. Athletes with cured prior HCV infection may be cleared for competition in all combat sports. Athletes with prior HBV infection and no detectable HBV DNA in blood can be cleared for competition in all combat sports. Athletes with latent HBV infection with detectable HVB DNA in blood have a small risk of disease reactivation, so they should not be cleared. Athletes with

HIV infection should be disqualified indefinitely regardless of current viral load or past/present antiretroviral therapy.

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PREAMBLE: DEVELOPMENT OF THIS STATEMENT

The Association of Ringside Physicians (ARP) is an international, non-profit organization dedicated to the health and safety of combat sports athletes. This Position Statement represents a collaborative effort among the ARP board of directors, emeritus board, and subject matter experts from the membership. It seeks to highlight issues in blood-borne infections unique to combat sports athletes.

The screening measures for blood-borne infections in combat sports athletes are critical to ensure the safety and health of all athletes. Blood-borne infections, such as HIV, HBV, and HCV, can be transmitted through contact with infected blood. Although there exists a small risk transmission during combat sports, minimizing this risk is of utmost importance to protect athletes.

An extensive literature search including but not restricted to Pubmed, Cochrane Reviews and non-indexed peer-reviewed articles published in online medical journals was performed to determine relevance of articles to the above blood-borne infections, combat sports, or sports in general. After review, discussion, and revision, the manuscript was formally approved by the authors and the entire board of directors as the official Position Statement of the ARP.

Hepatitis B

Epidemiology

There are an estimated 296 million carriers of HBV worldwide, with 1-5 million new infections per year, a prevalence of 3.5% and an estimated 820,000 annual mortality [1,2]. Since HBV is resistant to a spectrum of temperatures, as well as resistant to drying and alcohol, it can remain on surfaces for up to a week and has an increased risk of transmission when compared to HIV and HCV [3]. Moreover, when compared to HIV, HBV infection causes a higher viral load which can lead to an increased risk of transmission [3].

Transmission of HBV occurs through vertical transmission in pregnancy, sexual intercourse, mucus membrane exposure to infected blood or other body fluids, or percutaneously via contaminated needles [3,4]. Since HBV has a high risk of transmission, it is estimated that in sports, the risk of contracting HBV is one in every 850,000 to 4.25 million games [5]. It should be noted that this risk was estimated on extrapolating data from the national football league. There have been case reports of transmission of HBV in sports, however they are isolated cases and pertain to sumo wrestling, soccer and cross country sports [6,7,8]. There have been no reported cases of HBV transmission in sanctioned combat sports [3,5].

Pathophysiology

HBV is a partially double-stranded DNA virus belonging to the Hepadnaviridae family [9]. HBV is resistant to a wide range of temperatures and can even stay active on surfaces at 44°C for 7 days [10]. Within the double-

stranded DNA genome of HBV are 4 genes; the S, C, X and P genes which correspond to the surface envelope, core antigens, transcription proteins and polymerase proteins respectively. [11,13].

After reaching the host bloodstream HBV initially attaches to hepatocytes, the primary target cells in the liver, through interactions between its surface antigen (HBsAg) and host cell receptor [13,14]. After attachment, the virus enters the hepatocytes and undergoes DNA integration followed by viral protein transcription via covalently closed circular DNA (cccDNA) [15-18]. The cccDNA serves as a template for transcription of viral RNA, ensuring persistent infection and viral replication [15,17,18].

The host RNA polymerase transcribes viral messenger RNA to eventually encode viral proteins such as nucleocapsids which contain viral DNA. Once this occurs, HBV is released via exocytosis into the blood for further infection of other hepatocytes.

HBV induces both the innate and adaptive immune response. HBV elicits cellular lysis by CD4⁺ and CD8⁺ lymphocytes due to immune system recognition of HBsAg on infected cells [11]. Cytotoxic T lymphocytes (CTL), which are a part of the adaptive immune response, clears an HBV infection by killing infected cells and by producing antiviral cytokines such as IL-4, IL-13 and TNF- α [19,20]. During this process however, CTLs and inflammation from the cytokines induce hepatocellular damage.

During this acute phase, which lasts 1-6 months, the majority of patients are asymptomatic, but they can present with anorexia, nausea, vomiting, myalgia and icterus [13,21]. In both the acute and chronic phase, prolonged inflammation can lead to fibrosis, cirrhosis and hepatocellular carcinoma (HCC) [12,13]. Hepatitis D can also occur during a HBV infection as hepatitis D uses HBV surface antigen to infect hepatocytes [22]. Once the infection is eradicated, the prognosis is good for patients as less than 5 percent of adults progress from acute to chronic hepatitis B [23]. A chronic HBV infection is defined as persistence of HBsAg for more than six months and consists of 4 phases [13].

In the immune tolerant phase, there are high levels of HBV replication, as well as elevated HBeAg and HBV DNA. Majority of patients are asymptomatic during this phase due to immune tolerance and there is no active liver disease [16]. Immune-active is the next phase, which is characterized by HBeAg seroconversion and an increase in HBeAg clearance [25]. Inactive chronic HBV is the next phase. During this phase, patients are HBeAg negative and anti-HBe positive with undetectable HBV DNA [13,24]. During the last phase, immune-active with HBeAg negative, HBV replication increases which leads to active liver disease [26]. A chronic HBV infection can lead to cirrhosis, liver failure, or HCC. Additionally, HBV infection can be complicated by immune-mediated extrahepatic manifestations, such as polyarteritis nodosa, membranous nephropathy and membranoproliferative glomerulonephritis [27,28].

Diagnosis:

The diagnosis of HBV infection requires considering the probable stage of infection in the patient as well as differentiating immunity status. There are 3 main components of blood tests for HBV: surface antigen (HBsAg),

antibody against core antigen (Anti-HBc), and antibodies against surface antigen (Anti-HBs) [29,30]. The key finding that establishes a current HBV infection is positive HBsAg [31]. Testing positive for HBsAg or anti-HBc IgM is indicative of an acute infection. A positive result for over 6 months establishes a chronic infection [32]. During the chronic phase, both HBsAg and Anti-HBc IgG will be positive. Further testing is done to detect replication, transmissibility and viral load. HBeAg is a marker of HBV replication and transmissibility while HBV DNA quantifies viral load [29,31]; these are not required tests for combat sports athlete screening. An undetectable HBeAg with development of anti-HBe indicates that the patient has immunity or is an inactive carrier.

For the purposes of combat sports, the diagnosis of active HBV infection is made by a positive hepatitis B surface antigen, and this would preclude participation in combat sports in which blood is common and accepted due to active infectious status. A negative HBsAg essentially rules out active HBV infection [4-6,31].

	HBsAg	Anti-HBs	Anti-HBc	HBeAg	Anti-HBe	HBV DNA	Cleared for Competition
Immunity from vaccination	undetectable	+	undetectable	undetectable	undetectable	Not tested	Yes
Immunity from resolved infection	Undetectable	+	IgG+	Undetectable	Undetectable	Undetectable	Yes
Acute infection	+	Undetectable	IgM+	+	Undetectable	Undetectable	No
Chronic infection [Active]	+	Undetectable or +	IgG+	+	Undetectable	>2000 IU/mL	No
Chronic infection [Inactive]	+	Undetectable or +	IgG+	Undetectable	+	<2000 IU/mL	No

Table 1: Interpretation of Hepatitis B serology

Treatment

Acute HBV infection is self-limiting in more than 95% of immunocompetent adults and requires no treatment [31]. Therefore, patients that require pharmacological treatment are those with chronic or severe disease, with a focus on quality of life and improving survival. Currently the treatment goal is to achieve sustained suppression of HBV replication and liver inflammation, with the goal of preventing complications such as cirrhosis or HCC. Treatment has been documented to reverse liver fibrosis as well as reduce the risk of HCC [32]. Treatment

strategy includes nucleos(t)ide analog medications and/or pegylated interferon. The FDA approved medications for treatment include two formulations of interferon alpha (conventional and pegylated) and five nucleos(t)ide analogues (Lamivudine, Adefovir, Entecavir, Telbivudine, and Tenofovir) [32].

Prevention

In the United States, an estimated 580,000 to 2.4 million are living with HBV infection, with nearly 66% of patients not aware that they have contracted the infection [29]. Since infection is transmitted through contact with infected blood or body fluids, avoiding high risk activity such as multiple unprotected sexual encounters or sharing needles is imperative.

Prevention of HBV infection for the individual is done by avoiding transmission and inducing immunity through immunization. Due to the persistent transmission of HBV in the United States, the Centers for Disease Control and Prevention (CDC) published new recommendations for general screening in 2023 [29]. The rationale was to start with a universal screening test for adults older than 18 years old with a triple panel that includes surface antigen, surface antibody and core antibody results. This contrasts with previous recommendations that recommended only testing patients who were deemed high risk for infection. Including universal screening for all adults reduces ongoing transmission and is considered cost efficient to help prevent infection progression to cirrhosis, HCC and death. This also helps identify patients who no longer have detectable immunity to hepatitis B and may benefit from booster vaccination.

Recommendations for combat sports athletes to prevent infection are identical to other persons, including avoiding IV drug use, avoiding unprotected sexual encounters, and adhering to screening recommendations from the CDC. If the athlete does not have immunity, it is recommended for the athlete to obtain updated HBV vaccination. Activities in which the risk of exposure to blood is substantial should be prohibited for athletes with acute or chronic infection. It is this panel's expert opinion that testing for HBV should be done within 3 months of competition.

PROCEDURES FOR ATHLETES TESTING FOR HEPATITIS B

The various amounts of serological testing of HBV may make it difficult to interpret whether the combat athlete is safe to compete. The combat athlete should get tested with HBsAg, Anti-HBs, and Anti-HBc. The first step in determining if an athlete can compete is to determine the status of infection. An acutely infectious athlete will test positive for HBsAg and should not be cleared to compete. Athletes with negative HBsAg, Anti-HBc and Anti-HBs may compete, but are considered not immune and recommended to get updated immunization. Athletes with negative HBsAg, Anti-HBc and positive Anti-HBs are cleared to compete and are considered immune.

An athlete who has a history of a prior HBV infection will have different serologic test results. A typical example is an athlete that tests negative for HBsAg, but positive Anti-HBc and Anti-HBs. In this case, HBV DNA should be tested to detect viral material that can be transmitted through blood. Only those with undetectable HBV

DNA should be cleared to compete. These tests should be done within 3 months of each fight, and the decision to clear before each fight will rely on the HBV DNA being undetectable.

HIV

Epidemiology

Human Immunodeficiency Virus (HIV), which causes Acquired Immunodeficiency Syndrome (AIDS), was first identified in 1981. Since its discovery, it is estimated to have caused over 50 million fatalities globally [33]. In 2022 an estimated 39 million people were living with HIV, with an annual incidence of 1-3 million cases [34,35]. Of all cases globally, Sub-Saharan Africa accounts for more than 50% of cases [34]. Although the incidence of new cases has declined in twenty years, AIDS remains one of the top ten causes of death globally [36].

Transmission of HIV is primarily through sexual intercourse, sharing contaminated needles and via vertical transmission during pregnancy, childbirth and breastfeeding [37]. Although there is a risk of transmission of HIV in sports, with an increased risk with contact and collision sports, the risk of transmission is exceedingly low [38]. Transmission can occur from bleeding wounds or abrasions from an infected athlete to exposed mucous membranes of another athlete. The increased risk from contact sports is due to the higher risk of bleeding injuries and prolonged close contact [5].

It is estimated that the risk of contracting HIV in any sport is less than one in a million games [33, 5]. In regard to HIV, there have been no documented confirmed reports of HIV transmission during any sport, although it is still estimated that professional boxing and professional mixed martial arts carries the highest risk of transmission of HIV [5]. There have been no reported cases of HIV transmission in any sanctioned combat sports but there have been few case reports of HIV transmission during altercations [39-42].

Pathophysiology

HIV is a retrovirus that belongs to the family of Retroviridae, that primarily infects monocytes and macrophages [43,44]. Two types of HIV are known, HIV-1 and HIV-2, with the former having higher infectivity and virulence [45]. The HIV virus is composed of two copies of positive-sense single stranded RNA. Being a retrovirus, HIV integrates its own RNA into host cellular DNA via reverse transcriptase [44]. This process is error-prone, leading to genetic variability and the generation of diverse viral variants [46]. Upon activation, host RNA polymerase transcribes the viral DNA into messenger RNA (mRNA), which is then translated into viral proteins. The RNA genome of HIV includes structural proteins, regulatory proteins, its own reverse transcriptase, nucleocapsid proteins, proteases and several copies of p24, the capsid protein for HIV [44,47]. Once transcribed by the cellular host, HIV-RNA is packaged into p24 and eventually forms a mature, infectious virion [48]. Once the virus is released, it spreads throughout the body in the bloodstream and extracellular fluid until it encounters a CD4+ T cell or dendritic cell. The HIV virus then attaches to the host cell through interactions between its envelope glycoprotein (gp120) and CD4 as well as CXCR4 receptor molecules present on the surface of CD4+ T cells, as well as co-receptors such as CCR5 on dendritic cells [49,50]. Once attached, the HIV virus fuses with the host cell and the viral contents are released within the cell. The virus then continues replication.

The infected CD4+ cells then remain within lymph nodes for 2 - 4 weeks until HIV rapidly multiplies within the body due a poor host response to the virus. This marks the beginning of the acute phase of an HIV infection. During this phase, it is common for the patient to experience fever, fatigue, myalgia, rash and painful lymph nodes [47,51,52]. In this phase, CD4+ T cell numbers decrease due to cellular lysis and activation of CD8+ T cells which help kill infected CD4+ cells [53]. Plasma RNA levels decline due to the immune response, which takes weeks to months and marks the beginning of the latent phase [54]. In the latent phase, anti-HIV antibodies become detectable, indicating seroconversion and CD4+ count increases but will not go back to normal [55]. Over a period of months to years, the CD4+ cell count declines, effectively decreasing host immune response and HIV-RNA replication continues until the CD4+ count drops below 500 cells/microL. Between 500 cells/microL and 200 cells/mircoL, patients are prone to infections with oral candidiasis, herpes simplex or varicella zoster [56]. As the CD4+ count falls below 200 cells/microL; patients become highly susceptible to infections from Mycobacterium avium, Mycobacterium tuberculosis, Pneumocystis carinii, cytomegalovirus (CMV) and toxoplasmosis [55,56]. These infections, along with other conditions such as Kapsoi sarcoma and wasting syndrome, constitute AIDS-defining conditions. In order to be diagnosed with AIDS, a patient must have a CD4+ count less than 200 cells/microL or have an AIDS-defining condition [57,58]. The median survival time of patients with a CD4+ count of less than 50 cells/mircoL is 12 -18 months [59].

The initial symptoms of HIV are nonspecific and occur 2 - 4 weeks after exposure [5,34,35]. The most common are fever, lymphadenopathy, myalgia/arthritis, sore throat, GI symptoms and rash but none of them are specific to HIV [37,38]. The constellation of symptoms is known as acute retroviral syndrome and occur during the initial HIV infection [44,45]. During the latent stage, patients are usually asymptomatic. Before a patient's CD4 count decreases below 200 cells/microL, they will have nonspecific symptoms such as fatigue, night sweats, weight loss, fever and generalized lymphadenopathy [49,53]. More importantly, infections such as oral candidiasis, bacterial folliculitis, herpes simplex and varicella zoster may manifest [54,56,57]. Once the patient has less than 200 cells/microL or develops an AIDS-defining condition, the patient then has AIDS. At less than 200 cells/microL, the patient is at risk for Pneumocystic jirovecii, HSV, Progressive multifocal leukocencephalopathy and cryptosporidium [56-58]. At less than 100 cells/microL, there is a risk of developing cerebral toxoplasmosis, cryptococcal meningitis and disseminated CMV infections [49,53,56]. At less than 50 cells/microL, there is a risk of developing Mycobacterium avium complex [56-58].

Diagnosis

The diagnosis of HIV involves testing for both the antigen and antibody for HIV 1 and 2 via an immunoassay [60,62]. These tests are done as initial screening tests and can be done as either a combination of HIV antigen/antibody test (4th generation) or a HIV antibody ELISA [63-65]. The combination of the antigen/antibody test incorporates both HIV-1, HIV-2 and the p24 antigen. These will be positive 3 weeks to 3 months after first exposure. If positive, a confirmatory test is done via a HIV 1 and 2 antibody differentiation immunoassay [60,62]. If this test is positive, it confirms the diagnosis of HIV [64,64]. Newer 5th generation tests are able to distinguish an

infection with HIV-1 and HIV-2 [65,66]. If the test is negative or indeterminate, a nucleic acid amplification test [NAAT] is done which will determine the presence of HIV-RNA [60]. The benefit of a NAAT is that it can detect HIV earlier than any other test and can be done 10 days after suspected transmission [61]. It should be noted that patients that have a known exposure or potentially exposed should have a repeat antibody/antigen test done 4-6 weeks after exposure, and if negative then, repeated again 3 months after exposure [67,68].

There are no high-quality studies that give recommendations regarding the frequency of HIV testing for athletes in combat sports, and each state has varying recommendations on frequency of testing. Given that there exists a risk of transmission, it is recommended that an athlete be tested every 3 to 6 months, with a strong preference for every 3 months. In athletes that are high risk, we recommend that athletes be tested every three months. If the athlete has a known exposure to HIV, they should be disqualified from competition in sports where blood is common and accepted until they test negative for HIV at least 3 months after the exposure.

Treatment

The goal of HIV treatment is to disrupt the process of the virus multiplying and reduce the amount of virus in the body by targeting different stages of its life cycle [69-71]. Although HIV cannot be cured, the use of antiretroviral therapy [ART] can reduce transmission, reduce viral load, increase the CD4 count and improve the patient's quality of life [72-75].

There are six drug classes that encompass ART. Nucleoside Reverse Transcriptase Inhibitors (NRTIs), Non-Nucleoside Reverse Transcriptase Inhibitors (NNRTIs), Protease Inhibitors (PIs), Integrase Strand Transfer Inhibitors (INI), Fusion Inhibitors [FI] and Chemokine receptor antagonists[76-80]. Regardless of a patient's stage of HIV, all patients require treatment with ART. Although several ARTs exist, generally the treatment regime consists of 2NRTIs plus a NNRTI or 2NRTIs plus 1 PI or 2NRTIs plus INSTI or 1 NRTI plus 1 INI [Dolutegravir and lamivudine specifically] [81-86]. The selection of a treatment regime is dependent on drug toxicity, underlying illnesses, drug interactions, HIV resistance testing and access to care [84-86].

PROCEDURE FOR ATHLETES WHO TEST POSITIVE FOR HIV

Athletes with HIV infection (past positive HIV antigen/antibody test (4th generation) or HIV antibody ELISA, regardless of current viral load or past ART treatment) should be suspended indefinitely from competition in sports where blood is common and accepted, given the increased risk for viral transmission.

Hepatitis C

Epidemiology

There are an estimated 58 million chronic carriers of HCV, with 1-5 million new infections yearly and a prevalence of 0.7 percent [87,88]. It should be noted that these numbers are estimates due to lack of surveillance globally.

Transmission of HCV is primarily through use of contaminated needles. Other less common modes of transmission are through blood transfusions, sexual intercourse, body piercing and vertical transmission in pregnancy [89,90]. There have been no cases of HCV transmission through sports [91,92].

While no case reports or studies have linked sanctioned combat sports with HCV transmission, there are several case reports of persons without other risk factors or exposures acquiring HCV from bloody fights. These include a sheriff who had blood splashed into his eyes while breaking up a fight between two incarcerated individuals [93], a policeman whose hand was lacerated from punching a man in the mouth while making an arrest [94], and two family members who had a bloody fight and afterwards shared a handkerchief to wipe the blood off their bodies [95]. Given these documented cases, plus the significant proportion of HCV cases with no known exposure and one study showing contact sports to be a risk factor [96], it would appear there is an increased risk of HCV transmission between persons engaging in combative activities in which blood or body fluids are possibly exchanged.

Pathophysiology

HCV is a single-stranded positive-sense RNA virus belonging to the Flaviviridae family [97]. The genome of hepatitis C encodes three structural proteins, a core protein, two envelope proteins [E1 and E2], p7 and NS2 which are essential for viral production, and five nonstructural proteins that are required for replication [98-100]

HCV initially attaches to host cells, primarily hepatocytes in the liver, via mediation of glycosaminoglycans, CD81, claudin-1 and receptor B1 [101]. After attachment to the host cell, endocytosis occurs and the HCV positive-stranded RNA is uncoated which is then translated into a single polyprotein precursor. The polyprotein is then cleaved by both host and viral proteases into individual viral proteins and structural proteins [97,101]. These proteins are then transported to the host cell endoplasmic reticulum and form a replication complex with RNA dependent RNA polymerase [97,102,103]. The replication complex contains both a negative-sense and positive sense RNA [102,103]. Once viral genome replication occurs, it is packaged into the core and envelope proteins and it is released from the host cell. HCV continues to infect hepatocytes, which lasts 2 to 6 weeks until symptoms appear; however most patients are asymptomatic as symptoms occur in only 20% of patients [104].

Symptoms of HCV infection are nonspecific and include myalgia, fever, nausea and vomiting. Jaundice occurs only in 20% of patients [101]. For patients with an acute HCV infection, 50.8% of cases progress to chronic HCV infection [105]. A chronic infection is defined as persistent HCV-RNA for greater than 6 months [101,106]. Progression from an acute to a chronic infection is thought to be from an altered CD4+ T helper cell and cytotoxic T

cell response which causes necroinflammatory lesions in hepatocytes, however HCV is not cytopathic to cells. HCV employs various mechanisms to evade the host immune response, including rapid mutation rates, interference with innate immune signaling pathways, and modulation of host cell apoptosis [101].

Chronic inflammation then triggers fibrosis and eventually cirrhosis. The rate of fibrosis is increased in those with comorbid hepatitis B, HIV, alcohol use disorder and male sex [107]. Liver cirrhosis can then lead to ascites, spontaneous bacterial peritonitis, bleeding from esophageal varices, jaundice, and hepatic encephalopathy [97,103,104]. Extrahepatic complications of hepatitis C include insulin resistance, cardiomyopathy, membranoproliferative glomerulonephritis, porphyria cutanea tarda, mixed cryoglobulinemia and lichen planus [104,109-111]. Most importantly, approximately 50% of patients with chronic hepatitis C develop hepatocellular carcinoma, which carries a high mortality rate [112].

Diagnosis

Routine screening of HCV is recommended for all persons 18 years and older, according to recommendations from the CDC [113,114]. Justification for this approach includes HCV being a major source of morbidity and mortality, the high prevalence of asymptomatic infection, lack of a vaccine, and striving for lower transmission rates. In the general population, screening is recommended at least once in a lifetime, and periodically in persons with persistent risk factors [113]. These include IV drug use, prior blood transfusion from non-screened persons, sex with persons of known infection status, tattooing and body piercing, and incarceration [114-122]. Twenty to forty percent of patients infected with HCV do not have any risk factors and as a result, have acquired them by unknown means [115,123-127].

Screening for HCV infection involves a blood sample analyzed for the anti-HCV antibody using an immunoassay approved by the US Food and Drug Administration. Most samples require venous blood from phlebotomy, but point-of-care tests using capillary blood from fingerstick are also accurate [122,123]. A positive antibody test suggests either active HCV infection, past HCV infection, or rarely a false-positive with no past infection [113-115]. When anti-HCV is positive, a test for HCV-RNA should then be done, optimally with a “reflex” testing method where the same blood sample is automatically and immediately followed up with a HCV-RNA test [113,114]. An FDA-approved test with a detection level of ≤ 25 IU/ml should be used [114]. If the RNA test is positive, this confirms active infection [114]. If RNA is negative, then the patient is considered to have a cured past infection [115]. Since there is a delay of up to 3 months between time of infection and time of antibody seropositivity [126], initially negative results in persons with exposure or risk factors may need to be repeated in 3 months [127-129].

There are no studies to guide recommendations for timing and frequency of testing for HCV in combat sports athletes and each state varies in their recommendations [129]. In the presence of other risk factors for infection, however, infection can occur subsequent to a negative test, so repeated testing may be necessary to detect infection. Testing within 3 months of competition is recommended to detect recently acquired infections. Though

risk-based testing could reduce the testing burden, assessment of athletes' high-risk behaviors would be unreliable and inaccurate due to self-underreporting. Therefore, a more standardized screening interval is advised.

Treatment

Modern multi-drug direct action antiviral (DAA) therapy such as with ledipasiv/sofosbuvir, achieves cure in over 95% of persons with only 8 to 12 weeks of therapy [127]. DAA is safe and easy to administer. Retreatment may be needed in some high-risk persons such as those with cirrhosis or infection with genotype 3 virus [127]. Those with undetectable HCV-RNA 12 weeks after treatment are considered to have achieved sustained virologic response (SVR) and are considered cured, with the same follow-up and testing requirements as if they were never infected [127]. However, while anti-HCV antibodies usually persist lifelong, they do not protect from reinfection. Therefore, further testing should be based on risk factors for infection [114].

PROCEDURES FOR ATHLETES WHO TEST POSITIVE FOR HCV

Athletes with positive anti-HCV and HCV-RNA have active infection and may transmit the virus through blood or other body fluids, so they should be disqualified from competition in sports where blood is common and accepted. Athletes with positive anti-HCV but undetectable HCV-RNA are considered immune and have achieved SVR and may be cleared for all combat sports [129]. Future testing of these athletes should continue to include both anti-HCV and HCV-RNA.

Discussion

Testing for HIV, HBV and HCV is crucial for ensuring the safety of athletes in combat sports due to the higher risk of blood-to-blood contact. The authors, however, acknowledge the limited amounts of case reports describing transmission of these viruses in combat sports and the potential these sports carry for transmission. The statements formulated are based on data extrapolated from the literature, collective expertise, and review of current practices from each state commission and practicality.

Testing for HIV, HBV and HCV should be done for all combat sports athletes participating in any activity in which blood exposure is common and accepted due to the increased risk of transmission of these blood borne infections. Testing must be done via serum samples. Rapid tests are not acceptable means of testing.

The rationale for the 3 to 6 month testing window prior to competition is based on several factors. HIV, HBV and HCV can cause serious morbidity and increase the risk of mortality and can be lifelong infections, so prevention is paramount. Each virus also has a period of non-detectability between exposure and evidence of infection, sometimes up to 3 months. This time frame gives athletes and regulators a reasonable opportunity to secure the testing, submit documents, and act on results. Though testing within 3 to 6 months of competition imposes a significant burden upon athletes and regulators, it is necessary to preserve the integrity of combat sports while

protecting athletes from serious infections. Theoretically, a reduced frequency of testing for athletes without risk factors or known exposures would reduce the testing burden, but implementation of risk-factor-based screening is difficult or impossible. Therefore, the more frequent testing interval is recommended.

Athletes should optimally be tested for HIV, HBV and HCV within 3 months of competition, however within 6 months is acceptable. Testing within 3 to 6 months of competition is based on the pathophysiology of each virus and when serum levels are detectable. Discretion should be used for athletes with risk factors for contracting each virus, and the more frequent 3 months testing interval is recommended. For HIV, CD4+ lymphocytes start to decline 1 to 3 weeks after an infection. Conversely HIV-RNA increases during this time and reaches a peak at 6 weeks. Although HIV can be detected 10 days after initial infection, this would be impractical to low-risk individuals. HIV testing done at 3 months has been shown to be 99% accurate [37]. Testing should be done by a combination of antigen/antibody HIV-1/2 immunoassay (4th or 5th generation) followed by a confirmatory HIV-1/HIV-2 antibody differentiation immunoassay. If the antibody differentiation immunoassay is inconclusive then HIV-RNA testing needs to be done. Regarding HBV, serum with positive HBsAg, HBeAg and anti-HBc IgM levels indicate an acute infection and all of these peak at 3 months after infection in between the incubation phase and prodrome phase. During this time, it would be the best time to test athletes. Athletes should also be encouraged to be vaccinated against hepatitis B if they have not already done so.

For HCV, serum levels of anti-HCV peak at 2 - 3 months post infection. HCV should be tested via serum antibodies (anti-HCV). If positive, a follow-up PCR test to detect HCV-RNA is necessary to detect active infection. If a patient has been previously treated for HCV infection, then documentation of a completed treatment regimen and of SVR must be submitted. Confirmation of SVR requires negative serum anti-HCV testing at 12 weeks and 24 weeks after treatment.

Although it is recommended that fighters who test positive for HIV, Hepatitis B or C do not compete in combat sports, it is highly recommended that they continue to exercise. For patients with HIV, exercise has been shown to provide multiple benefits for patients living with HIV [130]. Exercise can enhance quality of life, cardiorespiratory fitness and mental health. Aerobic exercise especially, has been shown to improve in VO₂max, lean body mass, and depression symptoms however no significant changes in CD4 count and viral load occur [131,132]. Like HIV, patients with Hepatitis B and C are encouraged to exercise to increase cardiorespiratory fitness, reduce inflammation, reduce incidence of heart disease and diabetes. Although exercise does not reduce viral load, it can have a positive long-term impact on the patient [133].

Citations

1. The World Health Organization. Hepatitis B. <https://www.who.int/news-room/fact-sheets/detail/hepatitis-b> (Accessed on March 9th, 2024).
2. Ott JJ, Stevens GA, Groeger J, Wiersma ST. Global epidemiology of hepatitis B virus infection: new estimates of age-specific HBsAg seroprevalence and endemicity. *Vaccine*. 2012;30(12):2212-2219.
3. Beltrami EM, Williams IT, Shapiro CN, Chamberland ME. Risk and management of blood-borne infections in health care workers. *Clin Microbiol Rev*. 2000;13(3):385-407.
4. Kordi R, Wallace WA. Blood borne infections in sport: risks of transmission, methods of prevention, and recommendations for hepatitis B vaccination. *Br J Sports Med*. 2004;38(6):678-684; discussion 678-684.
5. McGrew CA. Blood-borne pathogens in sports. In: *and Practices of Primary Care Sports Medicine*, Garrett WE, Kirkendall DT, Squire DL (Eds), Lippincott Williams & Wilkins, Philadelphia 2001. P.247.
6. Kashiwagi S, Hayashi J, Ikematsu H, Nishigori S, Ishihara K, Kaji M. An outbreak of hepatitis B in members of a high school sumo wrestling club. *JAMA*. 1982;248(2):213-214.
7. Tobe K, Matsuura K, Ogura T, et al. Horizontal transmission of hepatitis B virus among players of an American football team. *Arch Intern Med*. 2000;160(16):2541-2545.
8. Gille G, Ringertz O, Zetterberg B. Serum hepatitis among Swedish track-finders. II. A clinical study. *Acta Med Scand*. 1967;182(2):129-135.
9. Lau JY, Wright TL. Molecular virology and pathogenesis of hepatitis B. *Lancet*. 1993 Nov 27. 342(8883):1335-40.
10. Pysopoulos NT. Hepatitis B. *Medscape*. October 20th, 2022. <https://emedicine.medscape.com/article/177632-overview#a3>

11. Tripathi N, Mousa OY. Hepatitis B. [Updated 2023 Jul 9]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK555945/>
12. Chisari FV, Isogawa M, Wieland SF. Pathogenesis of hepatitis B virus infection. *Pathol Biol (Paris)*. 2010 Aug;58(4):258-66.
13. Cecil RL. *Goldman's Cecil Medicine*. 24th ed. Acute Viral Hepatitis. 961-969 Elsevier/Saunders; 2012.
14. Yan H, Zhong G, Xu G, et al. Sodium taurocholate cotransporting polypeptide is a functional receptor for human hepatitis B and D virus. *Elife*. 2012;1:e00049.
15. Beck J, Nassal M. Hepatitis B virus replication. *World J Gastroenterol*. 2007;13(1):48-64.
16. Bruss V. Hepatitis B virus morphogenesis. *World J Gastroenterol*. 2007;13(1):65-73.
17. Wei L, Ploss A. Hepatitis B virus cccDNA is formed through distinct repair processes of each strand. *Nat Commun* 12, 1591 (2021). <https://doi.org/10.1038/s41467-021-21850-9>
18. Wei L, Ploss A. Mechanism of hepatitis b virus cccdna formation. *Viruses*. 2021;13(8):1463.
19. Ferrari C. HBV and the immune response. *Liver Int*. 2015 Jan;35 Suppl 1:121-8. doi: 10.1111/liv.12749. PMID: 25529097.
20. Iannacone M, Sitia G, Ruggeri ZM, Guidotti LG. HBV pathogenesis in animal models: recent advances on the role of platelets. *J Hepatol*. 2007;46(4):719-726.
21. Wright TL, Mamish D, Combs C, et al. Hepatitis B virus and apparent fulminant non-A, non-B hepatitis. *Lancet*. 1992;339(8799):952-955.
22. Taylor JM. Hepatitis delta virus. *Virology*. 2006;344(1):71-76.
23. Tassopoulos NC, Papaevangelou GJ, Sjogren MH, Roumeliotou-Karayannis A, Gerin JL, Purcell RH. Natural history of acute hepatitis B surface antigen-positive hepatitis in Greek adults. *Gastroenterology*. 1987;92(6):1844-1850.

24. Hui CK, Leung N, Yuen ST, et al. Natural history and disease progression in Chinese chronic hepatitis B patients in immune-tolerant phase. *Hepatology*. 2007;46(2):395-401.
25. Liaw YF, Chu CM, Lin DY, Sheen IS, Yang CY, Huang MJ. Age-specific prevalence and significance of hepatitis B e antigen and antibody in chronic hepatitis B virus infection in Taiwan: a comparison among asymptomatic carriers, chronic hepatitis, liver cirrhosis, and hepatocellular carcinoma. *J Med Virol*. 1984;13(4):385-391.
26. Bonino F, Rosina F, Rizzetto M, et al. Chronic hepatitis in HBsAg carriers with serum HBV-DNA and anti-HBe. *Gastroenterology*. 1986;90(5 Pt 1):1268-1273.
27. Guillevin L, Lhote F, Cohen P, et al. Polyarteritis nodosa related to hepatitis B virus. A prospective study with long-term observation of 41 patients. *Medicine (Baltimore)*. 1995;74(5):238-253.
28. Lai KN, Lai FM, Chan KW, Chow CB, Tong KL, Vallance-Owen J. The clinico-pathologic features of hepatitis B virus-associated glomerulonephritis. *Q J Med*. 1987;63(240):323-333.
29. Conners EE, Panagiotakopoulos L, Hofmeister MG, et al. Screening and Testing for Hepatitis B Virus Infection: CDC Recommendations — United States, 2023. *MMWR Recomm Rep* 2023;72(No. RR-1):1–25. DOI: <http://dx.doi.org/10.15585/mmwr.rr7201a1>
30. Dekker SE, Green EW, Ahn J. Treatment and Prevention of Acute Hepatitis B Virus. *Clin Liver Dis*. 2021 Nov;25(4):711-724. doi: 10.1016/j.cld.2021-06.002. Epub 2021 Sep 9. PMID: 34593149.
31. Trépo C, Chan HL, Lok A. Hepatitis B virus infection. *Lancet*. 2014 Dec 6;384(9959):2053-63. doi: 10.1016/S0140.6736(14)60220.8. Epub 2014 Jun 18. PMID: 24954675.
32. Nguyen MH, Wong G, Gane E, Kao JH, Dusheiko G. Hepatitis B Virus: Advances in Prevention, Diagnosis, and Therapy. *Clin Microbiol Rev*. 2020 Feb 26;33(2):e00046-19. doi: 10.1128/CMR.00046-19. PMID: 32102898; PMCID: PMC7048015.
33. World Health Organization Global Health Observatory Data <https://www.who.int/gho/hiv/en/> (Accessed on March 9th, 2024)

34. Joint United Nations Programme on HIV/AIDS. Global HIV & AIDS statistics — Fact sheet - 2022. <https://www.unaids.org/en/resources/fact-sheet> (Accessed March 9th, 2024)
35. <https://www.who.int/teams/global-hiv-hepatitis-and-stis-programmes/hiv/strategic-information/hiv-data-and-statistics> (Accessed March 9th, 2024)
36. GBD 2017 HIV collaborators. Global, regional, and national incidence, prevalence, and mortality of HIV, 1980, 2017, and forecasts to 2030, for 195 countries and territories: a systematic analysis for the Global Burden of Diseases, Injuries, and Risk Factors Study 2017. *Lancet HIV*. 2019;6(12):e831-e859.
37. "HIV and Its Transmission". U.S. Centers for Disease Control and Prevention (CDC). 2003.
38. McGrew C, MacCallum DS, Narducci D, et al. AMSSM position statement update: blood-borne pathogens in the context of sports participation. *Br J Sports Med*. 2020;54(4):200-207.
39. O'Farrell N, Tovey SJ, Morgan-Capner P. Transmission of HIV-1 infection after a fight. *Lancet*. 1992;339(8787):246.
40. Emerson CR, Quah SP. Transmission of HIV-1 infection due to a fist fight. *Int J STD AIDS*. 2008;19(2):131-132.
41. Ippolito G, Del Poggio P, Arici C, et al. Transmission of zidovudine-resistant HIV during a bloody fight. *JAMA*. 1994;272(6):433-434.
42. Abel S, Césaire R, Cales-Quist D, Béra O, Sobesky G, Cabié A. Occupational transmission of human immunodeficiency virus and hepatitis C virus after a punch. *Clin Infect Dis*. 2000;31(6):1494-1495.
43. International Committee on Taxonomy of Viruses (2002). "61. Retroviridae". National Institutes of Health. Archived from the original on October 2, 2006. Retrieved March 10th, 2024
44. Jonathan Weber, The pathogenesis of HIV-1 infection, *British Medical Bulletin*, Volume 58, Issue 1, September 2001, Pages 61–72, <https://doi.org/10.1093/bmb/58.1-61>
45. Naif HM. Pathogenesis of hiv infection. *Infect Dis Rep*. 2013;5(Suppl 1):e6.

46. Gilbert PB, McKeague IW, Eisen G, et al. Comparison of HIV-1 and HIV-2 infectivity from a prospective cohort study in Senegal. *Stat Med.* 2003;22(4):573-593.
47. Rossi E, Meuser ME, Cunanan CJ, Cocklin S. Structure, function, and interactions of the hiv-1 capsid protein. *Life (Basel).* 2021;11(2):100.
48. Moir S, Chun TW, Fauci AS. Pathogenic mechanisms of HIV disease. *Annu Rev Pathol.* 2011;6:223-248.
49. Swinkels HM, Justiz Vaillant AA, Nguyen AD, et al. HIV and AIDS. [Updated 2024 May 6]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK534860/>
50. Deng H, Liu R, Ellmeier W et al. Identification of a major co-receptor for primary isolates of HIV-1. *Nature*1996; 381: 661–6
51. Lackner AA, Lederman MM, Rodriguez B. HIV pathogenesis: the host. *Cold Spring Harb Perspect Med.* 2012;2(9):a007005.
52. Schacker T, Collier AC, Hughes J, Shea T, Corey L. Clinical and epidemiologic features of primary HIV infection. *Ann Intern Med.* 1996;125(4):257-264.
53. Brenchley JM, Schacker TW, Ruff LE, et al. CD4+ T cell depletion during all stages of HIV disease occurs predominantly in the gastrointestinal tract. *J Exp Med.* 2004;200(6):749-759.
54. Mellors J, Rinaldo C, Gupta P et al. Prognosis in HIV-1 infection predicted by the quantity of virus in plasma. *Science*1996; 272: 1167–70
55. Libman H. Pathogenesis, natural history, and classification of HIV infection. *Prim Care.* 1992;19(1):1-17.
56. Selik RM, Mokotoff ED, Branson B, et al. Revised Surveillance Case Definition for HIV Infection United States, 2014. *MMWR Recomm Rep* 2014; 63:1.

5.- Battistini Garcia SA, Guzman N. Acquired Immune Deficiency Syndrome CD4+ Count. [Updated 2023 Aug 14]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK513289/>

58. Justiz Vaillant AA, Gulick PG. HIV and AIDS Syndrome. [Updated 2022 Sep 20]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK534860/>

59. Yarchoan R, Venzon DJ, Pluda JM, et al. CD4 count and the risk for death in patients infected with HIV receiving antiretroviral therapy. *Ann Intern Med.* 1991;115(3):184-189.

60. Branson BM, Owen SM, Wesolowski LG, et al for Centers for Disease Control and Prevention and Association of Public Health Laboratories. Laboratory Testing for the Diagnosis of HIV Infection: Updated Recommendations. <http://stacks.cdc.gov/view/cdc/23447>. Published June 27, 2014.

61. Branson BM, Handsfield HH, Lampe MA, et al. Revised recommendations for HIV testing of adults, adolescents, and pregnant women in health-care settings. Center for disease control and prevention website. 2006. <https://www.cdc.gov/mmwr/preview/mmwrhtml/rr5514a1.htm> (Accessed August 16, 2017.)

62. Final recommendation Statement: Human Immunodeficiency Virus (HIV) Infection:Screening. US Preventive Task Force website. 2013.

63. Recommended laboratory HIV testing algorithm for serum or plasma specimens. Corporate Author(s) : National Center for HIV/AIDS, Viral Hepatitis, and TB. Prevention (U.S.). Division of HIV/AIDS Prevention.;Association of Public Health Laboratories.;Published Date : Updated January 2018. <https://stacks.cdc.gov/view/cdc/50872>

64. (U.S.); Association of Public Health Laboratories.; National Center for HIV/AIDS, Viral Hepatitis, and TB Prevention (U.S.). Division of HIV/AIDS Prevention. Published Date : 8/12/2016. URL : <https://stacks.cdc.gov/view/cdc/40790>

65. Use of the Determine HIV 1/2 Ag/Ab combo test with serum or plasma in the laboratory algorithm for HIV diagnosis. Centers for Disease Control and Prevention (U.S.);Association of Public Health Laboratories.; National Center for HIV/AIDS, Viral Hepatitis, and TB Prevention (U.S.). Division of HIV/AIDS Prevention.; Published Date : 10/4/2017. URL : <https://stacks.cdc.gov/view/cdc/48472>

- 6.- Robb ML, et al.; RV 217 Study Team. Prospective Study of Acute HIV-1 Infection in Adults in East Africa and Thailand. *N Engl J Med*. 2016 Jun 2;374(22):2120-30. doi: 10.1056/NEJMoa1508952. Epub 2016 May 18. PMID: 27192360; PMCID: PMC5111628.
67. Ridzon R et al. Simultaneous transmission of human immunodeficiency virus and hepatitis C virus from a needle-stick injury. *N Engl J Med*. 1997 Mar 27;336(13):919-22. doi: 10.1056/NEJM199703273361304. PMID: 9070472.
68. Kared H et al. HIV-specific regulatory T cells are associated with higher CD4 cell counts in primary infection. *AIDS*. 2008 Nov 30;22(18):2451-60. doi: 10-1097/QAD.0b013e328319edc0. PMID: 19005268; PMCID: PMC3195674.
69. Niu MT, Stein DS, Schnittman SM. Primary human immunodeficiency virus type 1 infection: review of pathogenesis and early treatment intervention in humans and animal retrovirus infections. *J Infect Dis*. 1993 Dec;168(6):1490-1501. doi: 10-1093/infdis/168.6.1490. PMID: 8245534.
70. Daar ES et al. Los Angeles County Primary HIV Infection Recruitment Network. Diagnosis of primary HIV-1 infection. Los Angeles County Primary HIV Infection Recruitment Network. *Ann Intern Med*. 2001 Jan 2;134(1):25-9. doi: 10-7326/0003-4819-134-1-20010102-.00010. PMID: 11187417.
71. Braun DL et al. Frequency and Spectrum of Unexpected Clinical Manifestations of Primary HIV-1 Infection. *Clin Infect Dis*. 2015 Sep 15;61(6):1013-21. doi: 10-1093/cid/civ398. Epub 2015 May 19. PMID: 25991469.
72. Crowell TA et al. RV254/SEARCH010 Study Group. Acute Retroviral Syndrome Is Associated With High Viral Burden, CD4 Depletion, and Immune Activation in Systemic and Tissue Compartments. *Clin Infect Dis*. 2018 May 2;66(10):1540-1549. doi: 10-1093/cid/cix1063. PMID: 29228130; PMCID: PMC5930255.
73. Gupta KK. Acute immunosuppression with HIV seroconversion. *N Engl J Med*. 1993 Jan 28;328(4):288-9. doi: 10-1056/NEJM199301283280419. PMID: 8380301.
74. Schacker T et al. Clinical and epidemiologic features of primary HIV infection. *Ann Intern Med*. 1996 Aug 15;125(4):257-64. doi: 10-7326/0003-4819-125-4-199608150-00001. Erratum in: *Ann Intern Med* 1997 Jan 15;126(2):174. PMID: 8678387.

75. Ambrosioni J et al. Hospital Clinic PHI Investigators. Neurological involvement in patients with acute/recent HIV-1 infection. A case-control study. *J Neurovirol*. 2017 Oct;23(5):679-685. doi: 10-1007/s13365-017-0548-6. Epub 2017 Jul 17. PMID: 28718069.

76. Andrade RM, Torriani FJ, Ellis RJ. Acute HIV infection presenting as fulminant meningoencephalitis with massive CSF viral replication. *Neurol Clin Pract*. 2014 Jun;4(3):256-259. doi: 10-1212/CPJ.0000000000000037. PMID: 25110623; PMCID: PMC4121464.

77. Hagberg L et al. Guillain-Barré syndrome as an early manifestation of HIV central nervous system infection. *Scand J Infect Dis*. 1986;18(6):591-2. doi: 10-3109/00365548609021668. PMID: 3468607.

78. Robb ML, et al.; RV 217 Study Team. Prospective Study of Acute HIV-1 Infection in Adults in East Africa and Thailand. *N Engl J Med*. 2016 Jun 2;374(22):2120-30. doi: 10-1056/NEJMoa1508952. Epub 2016 May 18. PMID: 27192360; PMCID: PMC5111628.

79. Cooper DA, Tindall B, Wilson EJ, Imrie AA, Penny R. Characterization of T lymphocyte responses during primary infection with human immunodeficiency virus. *J Infect Dis*. 1988 May;157(5):889-96. doi: 10-1093/infdis/157.5.889. PMID: 2966211.

80. <https://hivinfo.nih.gov/understanding-hiv/fact-sheets/hiv-treatment-basics>

81. Cohen MS, Chen YQ, McCauley M, et al. Prevention of HIV-1 Infection with Early Antiretroviral Therapy. *N Engl J Med Overseas Ed* 2011;365:493–505.

82. Cohen MS, Chen YQ, McCauley M, et al. Antiretroviral therapy for the prevention of HIV-1 transmission. *N Engl J Med Overseas Ed* 2016;375:830–9.

83. Trickey A, May MT, Vehreschild J-J, et al. Survival of HIV-positive patients starting antiretroviral therapy between 1996 and 2013: a collaborative analysis of cohort studies. *Lancet HIV* 2017;4:e349–e356

84. Eichner ER, Calabrese LH. Immunology and exercise. Physiology, pathophysiology, and implications for HIV infection. *Med Clin North Am* 1994;78:377–88.

85. Zanetti HR, da Cruz LG, Lourenço CL, et al. Does nonlinear resistance training reduce metabolic syndrome in people living with HIV? A randomized clinical trial. *J Sports Med Phys Fitness* 2017;57:678–84. 104
86. Hand GA, Phillips KD, Dudgeon WD, et al. Moderate intensity exercise training reverses functional aerobic impairment in HIV-infected individuals. *AIDS Care* 2008;20:1066–74
87. WHO Fact Sheet. Hepatitis C. Updated June 24, 2022. <https://www.who.int/news-room/fact-sheets/detail/hepatitis-c> (Accessed on March 9th, 2024).
88. Polaris Observatory HCV Collaborators. Global change in hepatitis C virus prevalence and cascade of care between 2015 and 2020: a modelling study. *Lancet Gastroenterol Hepatol.* 2022;7(5):396-415.
89. Centers for Disease Control and Prevention. Hepatitis C Surveillance 2020. <https://www.cdc.gov/hepatitis/statistics/2020surveillance/hepatitis-c.htm> (Accessed on September 21, 2022).
90. Murphy EL, Bryzman SM, Glynn SA, et al. Risk factors for hepatitis c virus infection in united states blood donors. Nhlbi retrovirus epidemiology donor study(Reds). *Hepatology.* 2000;31(3):756-762.
91. Karmochkine M, Carrat F, Dos Santos O, Cacoub P, Raguin G. A case-control study of risk factors for hepatitis C infection in patients with unexplained routes of infection. *J Viral Hepat.* 2006;13(11):775-782.
92. Mohd Suan MA, Said SM, Lim PY, Azman AZF, Abu Hassan MR. Risk factors for hepatitis C infection among adult patients in Kedah state, Malaysia: A case-control study. *PLoS One.* 2019;14(10):e0224459.
93. Rosen HR. Acquisition of hepatitis C by a conjunctival splash. *Am J Infect Control* 1997; 25:242-7.
94. Abel S, Cesaire R, Cales-Quist D, Bera O et al. Occupational transmission of human immunodeficiency virus and hepatitis C virus after a punch. *Clin Infect Dis* 2000; 31:494-5.
95. Bourliere M, Halfon P, Quentin Y, David P et al. Covert transmission of hepatitis C virus during bloody fisticuffs. *Gastroent* 2000; 119:507-11.
96. Karmochkine M, Carrat F, Dos Santons O, Cacoub P et al. A case-control study of risk factors for hepatitis C infection in patients with unexplained routes of infection. *J Viral Hepat* 2006; 13:775-82.

97. Basit H, Tyagi I, Koirala J. Hepatitis C. [Updated 2023 Mar 26]. In: StatPearls [Internet]. Treasure Island (FL): StatPearls Publishing; 2024 Jan-. Available from: <https://www.ncbi.nlm.nih.gov/books/NBK430897/>

98. Parigi TL, Torres MCP, Aghemo A. Upcoming direct acting antivirals for hepatitis C patients with a prior treatment failure. *Clin Mol Hepatol*. 2019 Dec;25(4):360-365.

99. Ghany MG, Liang TJ. Current and future therapies for hepatitis C virus infection. *N Engl J Med*. 2013 Aug 15. 369(7):679-80.

100. Dhawan V, Windle ML, Anand BS. Hepatitis C. *Medscape*. October 19th, 2019.

<https://emedicine.medscape.com/article/177792-overview#a3>

101. Cecil RL. *Goldman's Cecil Medicine*. 24th ed. Acute Viral Hepatitis. 963-969 Elsevier/Saunders; 2012.

102. Soi V, Daifi C, Yee J, Adams E. Pathophysiology and Treatment of Hepatitis B and C Infections in Patients With End-Stage Renal Disease. *Adv Chronic Kidney Dis*. 2019 Jan;26(1):41-50. doi: 10-1053/j.ackd.2018.10-004. PMID: 30876616.

103. Ranjith-Kumar CT, Kao CC. Biochemical Activities of the HCV NS5B RNA-Dependent RNA Polymerase. In: Tan SL, editor. *Hepatitis C Viruses: Genomes and Molecular Biology*. Norfolk (UK): Horizon Bioscience; 2006. Chapter 10.

Available from: <https://www.ncbi.nlm.nih.gov/books/NBK1629/>

104. Simoncini GM, Koren DE. Hepatitis C Update and Expanding the Role of Primary Care. *J Am Board Fam Med Hepatitis C Fact sheet*". WHO. 24 June 2022. Archived from the original on 31 January 2016.. 2019 May-Jun;32(3):428-430. doi: 10-3122/jabfm.2019.03.180286. PMID: 31068409.

105. "Hepatitis C Fact sheet". WHO. 24 June 2022. Archived from the original on 31 January 2016. Updated as required.

106. Wilkins T, Malcolm JK, Raina D, Schade RR. Hepatitis C: diagnosis and treatment. *Am Fam Physician*. 2010 Jun 1;81(11):1351-7. PMID: 20521755.

107. Ozaras R, Tahan V. Acute hepatitis C: prevention and treatment. *Expert Rev Anti Infect Ther*. 2009;7(3):351-361.

108. Dammacco F, Sansonno D. Therapy for hepatitis C virus-related cryoglobulinemic vasculitis. *N Engl J Med*. 2013 Sep 12;369(11):1035-45. doi: 10-1056/NEJMra1208642. PMID: 24024840.
109. Zignego AL, Ferri C, Pileri SA, Caini P, Bianchi FB; Italian Association of the Study of Liver Commission on Extrahepatic Manifestations of HCV infection. Extrahepatic manifestations of Hepatitis C Virus infection: a general overview and guidelines for a clinical approach. *Dig Liver Dis*. 2007 Jan;39(1):2-17. doi: 10-1016/j.dld.2006.06.008. Epub 2006 Aug 1. PMID: 16884964.
110. Matsumori A. Role of hepatitis C virus in cardiomyopathies. *Ernst Schering Res Found Workshop*. 2006;(55):99-120. doi: 10-1007/3-540-30822-9_7. PMID: 16329660.
111. Alter MJ. Epidemiology of hepatitis C virus infection. *World J Gastroenterol*. 2007 May 7;13(17):2436-41. doi: 10-3748/wjg.v13.i17.2436. PMID: 17552026; PMCID: PMC4146761.
112. Schillie S, Wester C, Osborne M, Wesolowski L et al. CDC recommendations for hepatitis C screening among adults – United States, 2020. *MMWR Recomm Rep* 2020; 69:1-17.
113. American Association for the Study of Liver Diseases (AASLD). Testing evaluation, and monitoring of hepatitis C. Updated 23 December 2023. https://www.hcvguidelines.org/sites/default/files/full-guidance-pdf/AASLD-IDS_A_HCVGuidance_December_19_2023.pdf. Accessed 2 Feb 2024.
114. Karmochkine M, Carrat F, Dos Santons O, Cacoub P et al. A case-control study of risk factors for hepatitis C infection in patients with unexplained routes of infection. *J Viral Hepat* 2006; 13:775-82.
115. Suan M, Said S, Lim P Azman et al. Risk factors for hepatitis C infection among adult patients in Kedah state, Malaysia: a case-control study. *PLoS One* 2019; 14:e0224459.
116. Kordi R, Neal K Pourfathollah AA, Mansournia MA et al. Risk of hepatitis B and C infections in Tehranian wrestlers. *J Athl Train* 2011;46:445-50.
117. Souto FJD, Da Silva AG, Yonamine F. Risk of hepatitis C among Brazilian ex-soccer players. *Mem Inst Oswaldo Crus* 2003; 98:1025-6.

118. Passos ADC, Figueiredo JFC. Hepatitis C among former athletes: association with the use of injectable stimulants in the past. *Mem Inst Oswaldo Cruz* 2008; 103:809-12.
119. Azevedo TCR, Filgueira NA, Lopes EP. Risk factors for hepatitis C virus infection in former Brazilian soccer players. *Epidemiol Infect* 2012; 140:70-3.
120. Aitken C, Delalande C, Stanton K. Pumping iron, risking infection? Exposure to hepatitis C, Hepatitis B and HIV among anabolic-androgenic steroid injectors in Victoria, Australia. *Drug Alcohol Depend* 2002; 65:303-8.
121. Rich JD, Dickinson BP, Pugatch D, Mylonakis E. The infectious complications of anabolic-androgenic steroid injection. *Int J Sports Med* 1999; 20:563-6.
122. Rosen HR. Acquisition of hepatitis C by a conjunctival splash. *Am J Infect Control* 1997; 25:242-7.
123. Abel S, Cesaire R, Cales-Quist D, Bera O et al. Occupational transmission of human immunodeficiency virus and hepatitis C virus after a punch. *Clin Infect Dis* 2000; 31:494-5.
124. Bourliere M, Halfon P, Quentin Y, David P et al. Covert transmission of hepatitis C virus during bloody fisticuffs. *Gastroent* 2000; 119:507-11.
125. Salari N, Darvishi N, Hemmaati M, Shohaimi S et al. Global prevalence of hepatitis C in prisoners: a comprehensive systematic review and meta-analysis. *Arch Virol* 2022; 167:1025-39.
126. Martinello M, Solomon SS, Terrault NA, Dore GJ. Hepatitis C. *Lancet* 2023; 402:1085-96.
127. McKenna O, Cunningham C, Gissane C, Blake C. Management of the extrahepatic symptoms of chronic hepatitis C: feasibility of a randomized controlled trial of exercise. *Am J Phys Med Rehabil* 2013; 92:504-12.
128. O’Gorman P, Strahan O, Ferguson D, Monaghan A et al. Improvement in cognitive impairment following a 12-week aerobic exercise intervention in individuals with non-cirrhotic chronic hepatitis C. *J Viral Hepat* 2021; 28:637-50.
129. McGrew C, MacCallum DS, Narducci D, Nuti R et al. AMSSM position statement update: blood-borne pathogens in the context of sports participation.

130. Garcia A, Fraga GA, Vieira RC, et al. Effects of combined exercise training on immunological, physical and biochemical parameters in individuals with HIV/AIDS. *J Sports Sci.* 2014;32(8):785-792.

131. Lox CL, McAuley E, Tucker RS. Aerobic and resistance exercise training effects on body composition, muscular strength, and cardiovascular fitness in an HIV-1 population. *Int J Behav Med.* 1996;3(1):55-69.

132. O'Brien KK, Tynan AM, Nixon SA, Glazier RH. Effectiveness of aerobic exercise for adults living with HIV: systematic review and meta-analysis using the Cochrane Collaboration protocol. *BMC Infect Dis.* 2016;16:182.

133. Ghany MG, Morgan TR, AASLD-IDSA Hepatitis C Guidance Panel. Hepatitis c guidance 2019 update: american association for the study of liver diseases-infectious diseases society of america recommendations for testing, managing, and treating hepatitis c virus infection. *Hepatology.* 2020;71(2):686-721.